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(71) Applicant: F. HOFFMANN-LA ROCHE AG
Postfach 3255
CH-4002 Basel(CH)

(72) Inventor: Hoshino, Tatsuo
Fueta 808-47
Kamakura-shi Kanagawa-ken(JP)
Inventor: Nomura, Setsuko
Matsuiso 202 Kameino 1015-2 Kameino
Fujisawa-shi Kanagawa-ken(JP)
Inventor: Sugisawa, Teruhide
Kohnandi 8-26 10, Kohnan-ku
Yokohama-shi Kanagawa-ken(JP)

(74) Representative: Aschert, Hubert et al
Grenzacherstrasse 124 Postfach 3255
CH-4002 Basel(CH)

(54) Fermentation process for producing 2-keto-L-gulonic acid.

(57) A process of producing 2-keto-L-gulonic acid by fermentative conversion of L-sorbose utilizing a fermentation system composed of component produced from a microorganism having the identifying characteristics of strain DSM No. 4025 and a yeast component.

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Fermentation process

The present invention relates to a process for producing 2-keto-L-gulonic acid by fermentative conversion of L-sorbose with a high yield.

2-Keto-L-gulonic acid is an important intermediate for the production of L-ascorbic acid into which it can be converted according to the well-known Reichstein method.

5 The fermentative production of 2-keto-L-gulonic acid from D-sorbitol or L-sorbose is known.

Thus, Japanese Patent Publication No. 40154/1976 discloses the production of 2-keto-L-gulonic acid from D-sorbitol by means of microorganisms of the genus *Acetobacter*, *Bacterium* or *Pseudomonas*, which are capable of oxidizing D-sorbitol under aerobic conditions, thus producing 2-keto-L-gulonic acid. The yield of this known process is, however, rather low, namely less than 6 g/l.

10 According to another known process, which is disclosed in "Acta Microbiologica Sinica" 21(2), 185-191, (1981), 2-keto-L-gulonic acid can be produced from L-sorbose by means of a mixed culture of microorganisms, which comprises *Pseudomonas striata* and *Gluconobacter oxydans*, the yield being 30 g/l when starting from a concentration of 70 g/l of L-sorbose, and 37 g/l when starting from a concentration of 100 g/l of L-sorbose.

15 Moreover, European Patent Publication No. 0221 707 discloses the production of 2-keto-L-gulonic acid from L-sorbose by means of *Pseudogluconobacter saccharoketogenes* with and without concomitant bacteria. However, the yield of this known process by means of *Pseudogluconobacter saccharoketogenes* per se is at most 55.3-87.6 g/l (conversion ratio: 34.2-54.1%) (See: page 13, Table 4 of European Patent Publication No. 0221 707).

20 Furthermore, European Patent Publication No. 0278447 discloses a process for the production of 2-keto-L-gulonic acid from L-sorbose by means of a mixed culture of microorganisms, one of which having the identifying characteristics of DSM No. 4025 and the other one having the identifying characteristics of DSM No. 4026 (a *Bacillus megaterium* strain). While the microorganism having the identifying characteristics of DSM No. 4025, particularly DSM No. 4025 itself, has, as such, substantially no ability to grow and
25 produce 2-keto-L-gulonic acid, the mixed culture, containing as the second component a *Bacillus megaterium* microorganism, does grow and does produce 2-keto-L-gulonic acid in yields of 40 g/l and more. Thus, the presence of the second microorganism component (*Bacillus megaterium*) was considered to be essential for the operability of the process disclosed in the European Patent Publication No. 0278447.

The present invention is based on the surprising finding, that it is possible to produce 2-keto-L-gulonic acid from L-sorbose using a microorganism having the identifying characteristics of strain DSM No. 4025, without concomitantly using another microorganism, i.e. *Bacillus megaterium*.
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The present invention is thus concerned with a process for producing 2-keto-L-gulonic acid by fermentative conversion of L-sorbose by the action of a fermentative system comprising:

- a) a pure culture of a microorganism having the identifying characteristics of microorganism DSM No. 4025 and having, as such, substantially no ability to produce 2-keto-L-gulonic acid from L-sorbose; and
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- b) yeast or a yeast product having the growth promoting activity of the yeast from which it is obtained.

According to the present invention, it is possible to produce 2-keto-L-gulonic acid from L-sorbose at a substantially improved yield, namely a yield of more than 72 g/l (conversion ratio: 93.7%) when starting
40 from a L-sorbose concentration of 80 g/l, and even more than 100 g/l (conversion ratio: 92.8%) when starting from a L-sorbose concentration of 100 g/l and at higher yields when starting from higher concentrations. Conversion ratio is calculated based on the amounts of 2-keto-L-gulonic acid produced and L-sorbose consumed.

The microorganisms used in the present invention have the identifying characteristics of strain DSM No. 4025 and has as such, substantially no ability to grow and to produce 2-keto-L-gulonic acid. The main identifying characteristics are:

negative oxidase test; ethanol is oxidized to acetic acid; D-glucose is oxidized to D-gluconic acid and 2-keto-D-gluconic acid; ketogenesis of polyalcohols; dihydroxyacetone is not substantially produced from glycerol; 2-keto-D-glucaric acid is produced from D-sorbitol and D-glucaric acid, but not from D-glucose, D-fructose, D-gluconic acid, D-mannitol or 2-keto-D-gluconic acid; polymorphic, apparently no flagella; brown pigment is produced from fructose; good growth when co-cultured in the presence of *Bacillus megaterium* or a cell extract thereof; streptomycin sensitive.
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In the present invention, a pure culture of the microorganism is used. The term "a pure culture of the microorganism" means a culture which contains the microorganism only, without any other concomitant microorganisms, such as *Pseudomonas striata* and *Bacillus megaterium*, thereby differing from the cultures

used in the processes disclosed in "Acta Microbiologica Sinica" 21 (2), 185-191 (1981) as mentioned above, and in European Patent Publication No. 278 447, where the use of mixed cultures is disclosed.

A specific and preferred microorganism strain has been deposited at the Deutsche Sammlung von Mikroorganismen in Göttingen under DSM No. 4025 on March 17, 1987 under the Budapest Treaty (the present address of this institute is: Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroder Weg 1b, D-3300 Braunschweig, Federal Republic of Germany).

Further preferred microorganisms are functional equivalents, subcultures, mutants and variants of the microorganism DSM No. 4025.

The cells of the DSM No. 4025 microorganism are rod-shaped with rounded ends. The diameter of a cell of the microorganism is, on an average about 0.3-0.6 μm , its length about 0.9-1.6 μm , mainly 1-1.5 μm .

In the preferable embodiment of the present invention, the production of 2-keto-L-gulonic acid is effected by cultivating the microorganism referred to above in a medium containing L-sorbose and yeast or a yeast product as well as appropriate nutrients. Alternatively, this process may be carried out by cultivating the microorganism in the above mentioned medium and, thereafter, bringing the whole cells collected from the culture or its cell-free extract into contact with L-sorbose. In carrying out the reaction of the present invention, any microorganism having the identifying characteristics of microorganism DSM No. 4025 can be utilized. Such microorganisms which have the identifying characteristics of strain DSM No. 4025 include subcultures, functional equivalents, variants and mutants thereof, which have the power to transform, in combination with yeast or a yeast product, L-sorbose into 2-keto-L-gulonic acid. Any conventional yeast or yeast product can be utilized. The yeast products which can be utilized in accordance with this invention can be any component obtained from the yeast which has the growth promoting properties of the yeast from which it is obtained.

The yeast used in the present invention includes any yeast belonging to the subclass Ascomycetes, particularly to the genus *Saccharomyces*, such as *Saccharomyces cerevisiae* (baker's yeast), *Saccharomyces carlsbergensis* (*Saccharomyces uvarum*) (brewer's yeast) or *Saccharomyces sake*, or to the genus *Schizosaccharomyces*, such as *Schizosaccharomyces pombe*, or to the genus *Pichia*, such as *Pichia membranaefaciens*, or to the genus *Hansenula*, such as *Hansenula anomala*, or yeast belonging to the subclass Hyphomycetes, particularly to the genus *Candida*, such as *Candida tropicalis* or *Candida utilis*, or to the genus *Torulopsis*, such as *Torulopsis versatilis* (*Candida versatilis*) or *Torulopsis holmii* (*Candida holmii*), or a yeast product thereof. The yeasts mentioned above are freely available from depositories or from commercial sources. Thus, the yeasts mentioned above belonging to the species *Saccharomyces cerevisiae* can be purchased from private companies for instance as baker's yeast. For example, the following baker's yeast can be purchased from the following companies:

- Oriental yeast (Oriental Yeast Co.)
- Kaneka yeast (Kanegafuchi Chemical Industry Co.)
- Daiya yeast (Kyowa Hakko Kogyo Co.)
- Sankyo yeast (Sankyo Co.)
- Chuetsu yeast (Chuetsu Yeast Co.)
- Yeast 45 (Toyo Jozo Co.)
- Nitten yeast (Nippon Beet-Sugar Co.)

Moreover, the following yeasts can be also purchased from IFO (Institute for Fermentation, Osaka, Japan),

- Saccharomyces cerevisiae* IFO 1234
- Saccharomyces carlsbergensis* IFO 0565
- (*Saccharomyces uvarum*)
- Saccharomyces sake* IFO 0309
- Candida tropicalis* IFO 1400
- Candida utilis* IFO 0396
- Torulopsis versatilis* IFO 10056
- (*Candida versatilis*)
- Torulopsis holmii* IFO 1629
- (*Candida holmii*)
- Schizosaccharomyces pombe* IFO 0362
- Pichia membranaefaciens* IFO 10215
- Hansenula anomala* IFO 10213

It is essential that the yeast used in the present process contains growth factors for the microorganisms used according to the present invention. The common properties of said growth factors are thermal stability and solubility in water.

The yeast or yeast products used in the present invention include so-called "fresh yeast", dry yeast, yeast extract and the like. The term "fresh yeast" herein means yeast cells just collected from the culture broth by means of centrifugation, filtration and the like. Baker's yeast as commercialized in conventional form is a specific representative of such "fresh yeast". However, fresh yeast, such as baker's yeast, after having been heated at e.g. 121°C, can also be used advantageously in the process of the present invention. As a matter of fact, the use of sterilized yeast is preferred in view of the fact that live yeast would metabolize part of the L-sorbose used as substrate and thereby reduce the 2-KGA yield. Thus, the use of sterilized yeast, such as sterilized baker's yeast, is a highly preferred embodiment of the present invention.

Where the microorganism is cultured in a nutrient medium containing L-sorbose as well as yeast or a yeast product, the microorganism is conveniently cultured in an aqueous medium under aerobic conditions.

The concentration of the above yeast according to the present invention is preferably from about 5 g/l to about 150 g/l by wet weight in the medium, preferably from about 20 g/l to about 100 g/l by wet weight in the medium.

The instant fermentation process may be carried out at a pH between about 4.0 and 9.0, preferably between about 6.0 and 8.0.

A preferred temperature range for carrying out the instant fermentation process is between about 13 and 36°C. More preferably, the instant fermentation process is carried out at a temperature between about 18 and 33°C.

While the fermentation period may vary, depending on pH, temperature and nutrient medium used, usually 2 to 5 days will bring about favourable results.

The concentration of the L-sorbose used as starting material in the instant process may vary between about 20 and 250 g/l, preferably between about 50 and about 200 g/l.

The culture medium used in the instant fermentation process usually contains such nutrients for the microorganisms as assimilable carbon sources, digestible nitrogen sources and inorganic substances, vitamins, trace elements and other growth promoting factors. In addition to L-sorbose used as starting material in the instant process, other substances which are carbon sources may also be added, such as glycerol, D-glucose, D-mannitol, D-fructose, D-arabitol and the like.

Various organic or inorganic substances may also be used as nitrogen sources in the instant process, such as meat-extract, peptone, casein, corn steep liquor, urea, amino acids, nitrates, ammonium salts and the like. As inorganic substances, magnesium sulfate, potassium phosphate, ferrous and ferric chlorides, calcium carbonate and the like may be used.

In the case where pregrown whole cells collected from the culture are used, the cultivation of the microorganism is carried out under the same or similar conditions as described above. These whole cells are used in an aqueous medium under aerobic conditions, no additional nutrients (in addition to the L-sorbose used as starting material) are necessary.

In case where cell-free extracts from the culture are used, these extracts are added to the substrate in an aqueous medium and are used in the conversion of L-sorbose to 2-keto-L-gulonic acid under aerobic conditions in a manner similar to that set forth above, no additional nutrients being necessary also in this case.

The 2-keto-L-gulonic acid obtained according to the present process can be isolated from the reaction mixture, e.g. by the formation of a salt or by using differences in properties between the product and the surrounding impurities, such as solubility, absorbability and distribution coefficient between the solvents. Adsorption, e.g. on ion exchange resins constitutes a convenient means for isolating the product. The thus obtained product may further be purified in a conventional manner, e.g. by recrystallization or chromatography.

Alternatively, the reaction mixture can be used directly for conversion to L-ascorbic acid by esterification, followed by enolization and lactonization.

The present invention is illustrated by the following Examples:

Example 1

A seed culture medium containing L-sorbose 8.0% (separately sterilized), glycerol 0.05%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 1.75%, yeast extract B-II (Oriental Yeast Co.) 0.25%, CaCO_3 0.5% and urea 0.5% (separately sterilized) (pH 7.0 before sterilization) was distributed into test tube (5 ml each) and sterilized at 121°C for 20 minutes. Into this seed culture medium, one loopful amount of the cells of microorganism DSM No. 4025 grown on a slant culture medium containing glycerol 3.0%, $\text{MgSO}_4 \cdot 0.25\%$,

corn steep liquor 1.75% yeast extracts B-II 0.25%, CaCO_3 0.5%, urea 0.5% (separately sterilized) and agar 2.0% (pH 7.0 before sterilization) at 27 °C for four days was inoculated and incubated at 30 °C for 24 hours. The resulting seed culture was inoculated into 50 ml of the same seed culture medium as described above in a 500 ml Erlenmeyer flask and incubated for 24 hours at 30 °C. The basal production medium (50 ml) containing L-sorbose 8.0% (sterilized separately), glycerol 0.05%, urea 1.0% (sterilized separately), $\text{MgSO}_4 \cdot 7 \text{H}_2\text{O}$ 0.25%, CaCO_3 1.5% and corn steep liquor 3.0% was supplemented with yeast extract B-II 0.5% and/or baker's yeast (Oriental Yeast Co.) 5% in a 500 ml Erlenmeyer flask and sterilized at 121 °C for 20 minutes. Into these production media, 5 ml each of the seed culture as prepared above was added and incubated at 30 °C for 4 days at an agitation speed of 180 rpm.

By high performance liquid chromatographic analysis, 2-keto-L-gulonic acid (2-KGA) content in the cultured broth supplemented with baker's yeast was determined to be 72.1 g/l (conversion rate: 93.7%). On the other hand, 2-KGA content of the broths supplemented with yeast extract B-II and yeast extract B-II/baker's yeast were 57.1 (conversion rate: 88.8%) and 64.9 g/l (conversion rate: 94.4%), respectively.

Example 2

A seed culture medium containing L-sorbose 8.0% (separately sterilized), glycerol 0.05%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 1.75%, baker's yeast 5.0%, CaCO_3 0.5% and urea 0.5% (separately sterilized) (pH 7.0 before sterilization) was distributed into test tubes (5 ml each) and sterilized at 121 °C for 20 minutes. Into this seed culture medium, one loopful amount of the cells of microorganism DSM No. 4025 grown on a slant culture medium containing D-mannitol 5.0%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 1.75%, baker's yeast 0.25%, CaCO_3 0.5%, urea 0.5% (separately sterilized) and agar 2.0%, (pH 7.0 before sterilization) at 27 °C for four days was inoculated and incubated at 30 °C for 24 hours. The resulting seed culture was inoculated into 50 ml of the same seed culture medium as described above in a 500 ml Erlenmeyer flask and incubated for 24 hours at 30 °C. The basal production medium (50 ml) containing L-sorbose 10.0% (sterilized separately), glycerol 0.05%, urea 1.6% (sterilized separately), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, CaCO_3 1.5% and corn steep liquor 3.0% was supplemented with the yeasts as listed in Table 1 in 500 ml Erlenmeyer flasks and sterilized for 20 minutes at 121 °C. Into these production media, 5 ml each of the seed culture as prepared above was added and incubated at 30 °C for 4 days with the agitation speed of 180 rpm.

2-KGA values of flasks of various conditions are shown in Table 1.

Table 1

Effect of various yeasts used as medium component on 2-KGA production					
Yeasts	Source	Amount %	2-KGA (g/l)	Remaining L-sorbose (g/l)	Cell growth
Yeast extract					
B-II	Oriental	0.25	68.5	31.1	+
		0.5	74.1	25.3	+
Yeast powder					
HG (dry yeast)	Oriental	0.25	75.6	23.0	+
		0.5	79.4	20.9	+
Yeast 20A (Dry yeast)	Bovril ⁽¹⁾	0.25	84.2	15.3	++
		0.5	87.3	12.0	++
Saf-instant (dry yeast)	S.I. Lesaffre ⁽²⁾	0.25	81.1	17.4	++
		0.5	86.0	13.8	++
Oriental dry yeast	Oriental	0.25	76.8	21.3	++
		0.5	86.5	13.7	++
Baker's yeast (S. cerevisiae)	Oriental	5.0	93.5	8.9	+++
		6.25	97.2	4.4	+++
None		0	4.0	93.0	±
Degree of cell growth: ± bad → +++ good					

(1) *Saccharomyces cerevisiae* available from Oriental Yeast Co.

(2) Baker's yeast

Example 3

In the same manner as described in Example 2, the seed culture was prepared, inoculated into 50 ml of the production medium containing L-sorbose 9.0% (separately sterilized), glycerol 0.05%, corn steep liquor 3.0%, urea 1.4% (separately sterilized), baker's yeast 5.6%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25% and CaCO_3 1.5% in a 500 ml Erlenmeyer flask and incubated at 30 °C for 4 days at an agitation speed of 180 rpm.

As a result, 89.0 g/l (conversion rate: 91.8%) of 2-KGA was produced.

Example 4

In the same manner as described in Example 2, the seed culture was prepared, inoculated into 50 ml of the production medium containing L-sorbose 10.0% (separately sterilized), glycerol 0.05%, corn steep liquor 3.0%, urea 1.6% (separately sterilized), baker's yeast 6.25%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25% and CaCO_3 1.5% in a 500 ml Erlenmeyer flask and incubated at 30 °C for 4 days at an agitation speed of 180 rpm.

As a result, 97.2 g/l (conversion rate: 90.2%) of 2-KGA was produced.

Example 5

In the same manner as described in Example 2, the seed culture was prepared, inoculated into 50 ml of

the production medium containing L-sorbose 12.0% (separately sterilized), glycerol 0.05%, corn steep liquor 3.0%, urea 1.9% (separately sterilized), baker's yeast 7.5%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25% and CaCO_3 1.5% in a 500 ml Erlenmeyer flask and incubated at 30 °C for 4 days at an agitation speed of 180 rpm. As a result, 98.5 g/l (conversion rate: 89.4%) of 2-KGA was produced.

Example 6

In the same manner as described in Example 2, the seed culture was prepared, inoculated into 50 ml of the production medium containing L-sorbose 8.0% (separately sterilized), glycerol 0.05%, corn steep liquor 3.0%, urea 1.25% (separately sterilized), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, CaCO_3 1.5% and 2 to 5% of baker's yeast, and incubated at 30 °C for 3 to 4 days.

As shown in Table 2, 82.6 g/l (conversion rate: 95.8%) of 2-KGA was produced in 4 days incubation in the medium supplemented with 4% of baker's yeast.

Table 2

Effect of concentration of baker's yeast on 2-KGA production			
Baker's yeast	2-KGA produced (g/l)		(Conversion: %)
	3 days	4 days	
2	75.1	77.5	(95.4)
3	80.6	81.3	(94.3)
4	81.6	82.6	(95.8)
5	81.8	82.6	(95.8)

Example 7

In the same manner as described in Example 2, 200 ml of the seed culture was prepared. Into a 3 l jar fermentor, 1.2 l of basal medium containing glycerol 0.05%, corn steep liquor 3.0%, baker's yeast 5.0% and $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25% to be made up to 2.0 l, was added and sterilized at 121 °C for 20 minutes. L-sorbose 8% (final concentration) and urea 1.25% (final concentration) were separately sterilized and added into the fermentor. After the inoculation by the seed culture, the culture volume was adjusted to 2 liters by addition of sterilized water. The fermentation was carried out at 30 °C, 800 rpm for agitation and 1 liter/minute for aeration. The pH of the culture was maintained at 7.0 by 4N of Na_2CO_3 .

In 40 hours fermentation, 84 g/l (conversion rate: 97.4%) of 2-KGA was produced.

Example 8

In the same manner as described in Example 7, 3 l jar fermentation was carried out in a medium containing L-sorbose 10.0%, urea 1.6%, glycerol 0.05%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 3.0% and baker's yeast 6.25%.

As a result, 100.0 g/l (conversion rate: 92.8%) of 2-KGA was produced in 50-hours fermentation.

Example 9

In the same manner as described in Example 7, 3 l jar fermentation was carried out in a medium containing L-sorbose 12.0%, urea 1.9%, glycerol 0.05%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 3.0% and baker's yeast 7.5%.

As a result, 98.0 g/l (conversion rate: 92.3%) of 2-KGA was produced in 75-hours fermentation. 21.5 g/l of L-sorbose remained unconsumed.

Example 10

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A 3 l jar fermentation was started in the same manner as described in Example 7. On the other hand, 80 g of L-sorbose was solubilized in 400 ml of water and sterilized at 121°C for 20 minutes and fed continuously into the fermentor at a feeding rate of about 11 ml/min from 6 to 42 hours of fermentation. After the termination of L-sorbose feeding, the fermentation was continued for further 25 hours. The total fermentation period was 67 hours. In the cultured broth (2.6 l), 95 g/l of 2-KGA was produced. Totally, 247 g of 2-KGA was produced from 240 g of L-sorbose. Molar conversion rate: 95.5%.

Example 11

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A 3 l jar fermentation was started in the same manner as described in Example 7. On the other hand, 120 g L-sorbose was solubilized in 400 ml of water and sterilized at 121°C for 20 minutes and fed continuously into the fermentor at a feeding rate of about 11 ml/min from 6 hours to 42 hours of fermentation. After the termination of L-sorbose feeding, the fermentation was continued for further 35 hours. The total fermentation period was 77 hours. In the cultured broth (2.7 l), 104 g/l of 2-KGA was produced. Totally, 280.8 g of 2-KGA was produced from 280 g of L-sorbose. Molar conversion rate: 93.1%.

Example 12

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A 3 l jar fermentation was started in the same manner as described in Example 7. On the other hand, 160 g of L-sorbose was solubilized in 400 ml of water and sterilized at 121°C for 20 minutes and fed continuously into the fermentor at a feeding rate of about 11 ml/min from 6 hours to 42 hours fermentation continuously. After the termination of L-sorbose feeding, the fermentation was continued for further 49 hours. The total fermentation period was 91 hours. In the cultured broth (2.7 l), 110 g/l of 2-KGA was produced. Totally, 297 g of 2-KGA was produced from 320 g of L-sorbose. Molar conversion rate: 86.1%.

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Example 13

In the same manner as described in Example 2, the seed culture was prepared. 5 ml each of the seed culture was inoculated into 50 ml of the production media containing L-sorbose 10% (sterilized separately), glycerol 0.05%, urea 1.6% (sterilized separately), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, corn steep liquor 3.0%, CaCO_3 1.5% and baker's yeast from various suppliers as listed in Table 3, 6.25% in 500 ml of Erlenmeyer flask. The fermentation was carried out at 30°C for 4 days at an agitation speed of 180 rpm. The results are shown in Table 3.

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Table 3

Effect of various baker's yeasts on 2-KGA production			
Type of baker's yeast	2-KGA (g/l)	(Conversion) (%)	Remaining L-sorbose (g/l)
Oriental yeast	97.2	(94.4)	4.4
Kaneka yeast	91.4	(90.5)	6.3
Daiya yeast	94.6	(92.3)	4.9
Sankyo yeast	87.4	(88.7)	8.6
Chuhetsu yeast	93.7	(92.2)	5.7
Yeast 45	90.1	(89.5)	6.6
Nitten yeast	97.2	(93.8)	3.8

Example 14

The basal production medium (5 ml), which contained L-sorbose 8% (sterilized separately), glycerol 0.05%, urea 1.25% (sterilized separately), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, CaCO_3 1.5% and corn steep liquor 3.0%, (pH 7.5 before sterilization), was supplemented with yeast cells (5% wet weight/volume), distributed in test tubes (1.8 x 20 cm) and sterilized at 121 °C for 20 minutes. As the yeast cells cultures of *Saccharomyces sake* IFO 0309, *Schizosaccharomyces pombe* IFO 0362, *Saccharomyces carlsbergensis* IFO 0565, *Saccharomyces cerevisiae* IFO 1234, *Candida utilis* IFO 0396, *Candida tropicalis* IFO 1400, *Torulopsis holmii* IFO 1629, *Torulopsis versatilis* IFO 10056, *Hansenula anomala* IFO 10213 and *Pichia membranaefaciens* IFO 10215 on an agar culture medium containing malt extract (Difco) 1.0%, yeast extract (Difco) 0.1%, soytone (Difco) 0.1%, glucose 1.0% and agar 2.0%, grown at 27 °C for 4 days were used. Into the production media, 0.5 ml each of the seed culture of microorganism DSM No. 4025 as prepared in the same manner as described in Example 2 was inoculated and incubated at 30 °C for 4 days.

2-KGA values of tubes in various conditions are shown in Table 4.

Table 4

Effect of the yeast belonging to genus <i>Saccharomyces</i> , <i>Candida</i> , <i>Torulopsis</i> , <i>Hansenula</i> and <i>Pichia</i> used as medium component on 2-KGA production			
Yeasts	amount (%)	2-KGA (g/l)	Remaining L-sorbose (g/l)
<i>Saccharomyces sake</i> IFO 0309	5.0	74.37	5.56
<i>Schizosaccharomyces pombe</i> IFO 0362	5.0	77.99	1.46
<i>Saccharomyces carlsbergensis</i> IFO 0565	5.0	70.82	2.61
<i>Saccharomyces cerevisiae</i> IFO 1234	5.0	72.39	3.89
<i>Candida utilis</i> IFO 0396	5.0	71.05	0
<i>Candida tropicalis</i> IFO 1400	5.0	80.95	2.91
<i>Torulopsis holmii</i> IFO 1629	5.0	72.20	6.32
<i>Torulopsis versatilis</i> IFO 10056	5.0	78.01	0
<i>Hansenula anomala</i> IFO 10213	5.0	75.91	1.97
<i>Pichia membranaefaciens</i> IFO 10215	5.0	65.56	12.78

Referential Example

A seed culture medium S1 containing L-sorbose 8.0% (separately sterilized), glycerol 0.05%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, yeast extract B-II 1.5% and CaCO_3 1.5% (separately sterilized) (pH 7.0 before sterilization) was distributed into test tubes (5 ml each) and sterilized at 121 °C for 20 minutes. Into this seed culture medium, one loopful amount of the cells of *Gluconobacter oxydans* U-13 (FERM-BP No. 1269) grown on the slant culture medium was inoculated and incubated at 30 °C for 48 hours. The resulting seed culture was inoculated into 50 ml of the same seed culture medium as described above in a 500 ml Erlenmeyer flask and incubated for 48 hours at 30 °C. On the other hand, another seed culture was prepared using the seed culture medium S2 containing L-sorbose 8.0%, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.25%, glycerol 0.05%, CaCO_3 1.5%, urea 0.5%, corn steep liquor 1.75% and baker's yeast 5.0% in the same manner as described above. The seed cultures S1 and S2, thus prepared were inoculated into the production medium in the same manner as described in Example 2. The fermentation was carried out at 30 °C for 4 days.

The 2-KGA yield obtained was 12.8 g/l in the case of culture medium S1 and 10.2 g/l in the case of culture medium S2. This shows that *Gluconobacter oxydans* U-13 (FERM-BP No. 1269) which, as a pure culture, has ability to grow and produce 2-KGA, did not give 2-KGA in high yield in yeast-containing production media.

Claims

1. Process for producing 2-keto-L-gulonic acid by fermentative conversion of L-sorbose by the action of a fermentative system comprising:
 - a) a pure culture of a microorganism having the identifying characteristics of microorganism DSM No. 4025 and having, as such, substantially no ability to produce 2-keto-L-gulonic acid from L-sorbose; and
 - b) yeast or a yeast product having the growth promoting activity of the yeast from which it is obtained.
2. Process according to claim 1, characterized in that a microorganism corresponding to DSM No. 4025, a functional equivalent, subculture, mutant or variant thereof is used.
3. Process according to claim 1 or 2, characterized in that yeast belonging to the subclass Ascomycetes, particularly to the genus *Saccharomyces*, such as *Saccharomyces cerevisiae* (baker's yeast), *Saccharomyces carlsbergensis* (*Saccharomyces uvarum*) (brewer's yeast) or *Saccharomyces sake*, or to the genus *Schizosaccharomyces*, such as *Schizosaccharomyces pombe*, or to the genus *Pichia*, such as *Pichia membranaefaciens*, or to the genus *Hansenula*, such as *Hansenula anomala*, or yeast belonging to the subclass Hyphomycetes, particularly to the genus *Candida*, such as *Candida tropicalis* or *Candida utilis*, or to the genus *Torulopsis*, such as *Torulopsis versatilis* (*Candida versatilis*) or *Torulopsis holmii* (*Candida holmii*), or a yeast product thereof is used as medium component.
4. Process according to claim 1, 2 or 3, characterized in that fresh yeast is used as medium component, preferably after sterilisation.
5. A process according to claim 1, wherein the yeast is used at a concentration of from about 5 g/l to about 150 g/l by wet weight, preferably from about 20 g/l to about 100 g/l by wet weight.
6. A process according to claim 1, wherein L-sorbose is used at a concentration of from about 20 g/l to about 250 g/l, preferably from about 50 g/l to about 200 g/l.
7. A process according to any one of claims 1-6, wherein 2-keto-L-gulonic acid is produced in a yield of at least 72 g/l, preferably at least 100 g/l.
8. A process according to any one of claims 1-7, wherein the fermentation is carried out at pH between about 4.0 and 9.0, preferably between about 6.0 and 8.0.
9. A process according to any one of claims 1-8, wherein the fermentation is carried out at a temperature between about 13 and 36 °C, preferably at between about 18 and 33 °C.



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EUROPÄISCHER RECHERCHENBERICHT

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EINSCHLÄGIGE DOKUMENTE			
Kategorie	Kennzeichnung des Dokuments mit Angabe, soweit erforderlich, der maßgeblichen Teile	Betrifft Anspruch	KLASSIFIKATION DER ANMELDUNG (Int. Cl.5)
D,A	EP-A-0 278 447 (INSTITUTE OF MICROBIOLOGY ACADEMIA SINICA) -----		C 12 P 7/60 C 12 P 39/00
			RECHERCHIERTE SACHGEBIETE (Int. Cl.5)
			C 12 P
Der vorliegende Recherchenbericht wurde für alle Patentansprüche erstellt			
Recherchenort DEN HAAG		Abschlußdatum der Recherche 22-12-1989	Prüfer DESCAMPS J.A.
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